



Low-Cost Water Quality Testing Methods

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MARCH 21 2016

INTRODUCTION

- ▶ Tyler Druhot
- ▶ Arjun Bir

BIG PICTURE

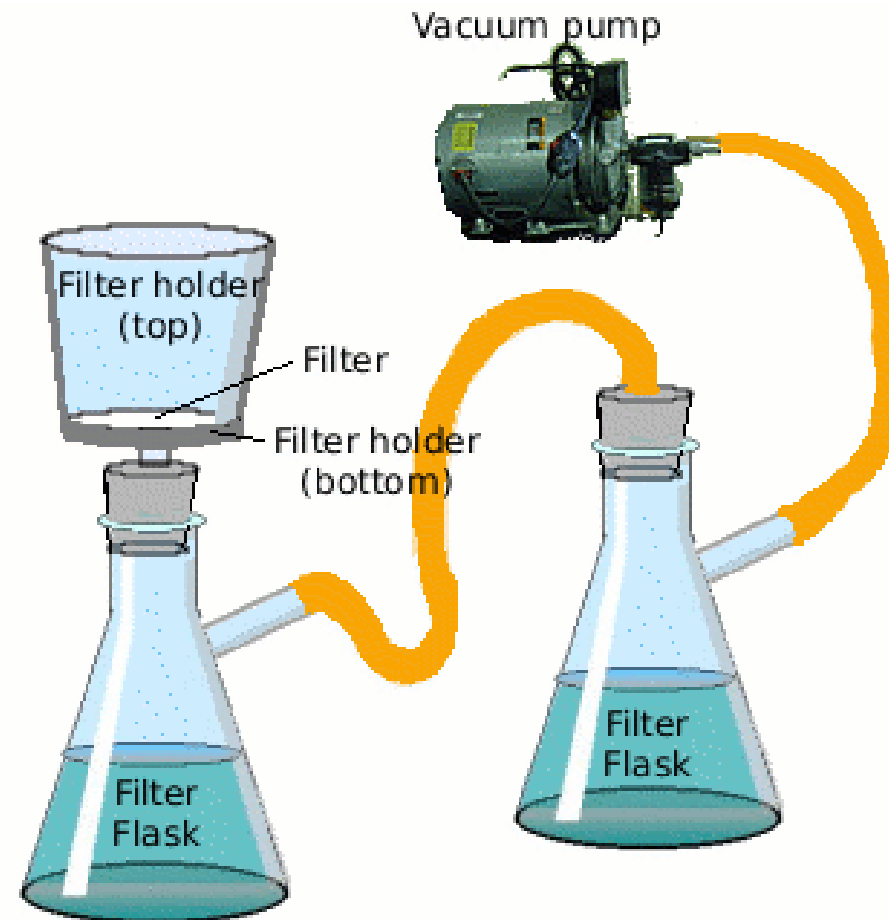
- ▶ Sustainable Development Goals: Access to safe drinking water for all by 2030
 - ▶ Need for scalable methods – microbial quality metrics being considered by UNICEF
- ▶ For all our partners present

CURRENT METHOD: CHALLENGES AND LIMITATIONS

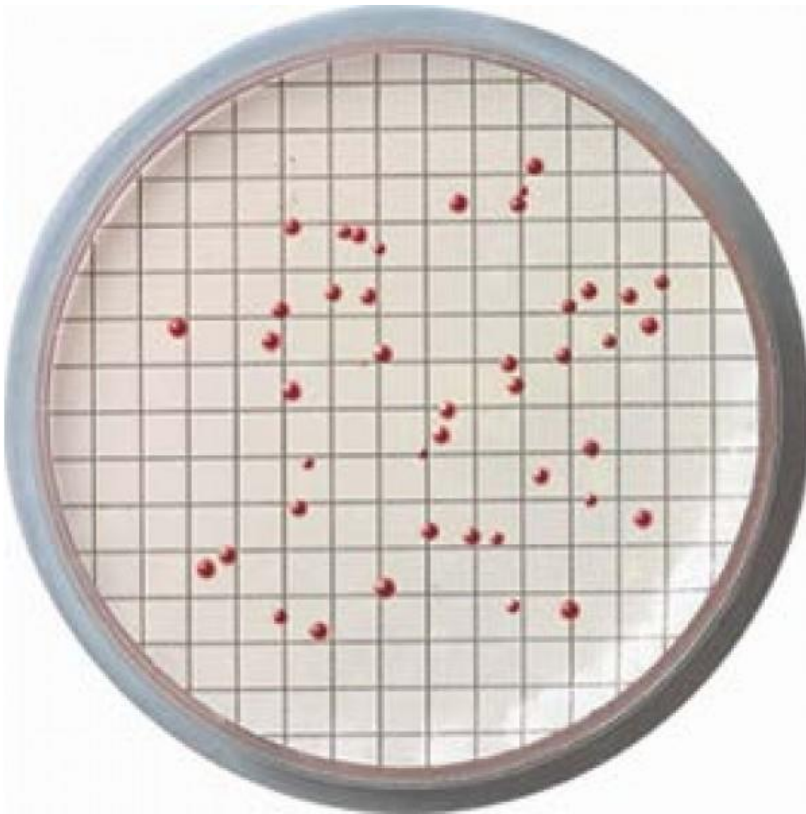
MEMBRANE FILTRATION

- ▶ Gold standard method
- ▶ Data output: *E. coli* Count (CFU/100mL)

MEMBRANE FILTRATION



MEMBRANE FILTRATION



- ▶ WHO Risk Levels:
 - ▶ Safe: <1 CFU/100mL
 - ▶ Low risk: 1-10 CFU/100mL
 - ▶ Medium risk: 1-100 CFU/100mL
 - ▶ High risk: 101+ CFU/100mL

MEMBRANE FILTRATION

PROS

- ▶ Very accurate
- ▶ Well characterized
- ▶ Widely used

CONS

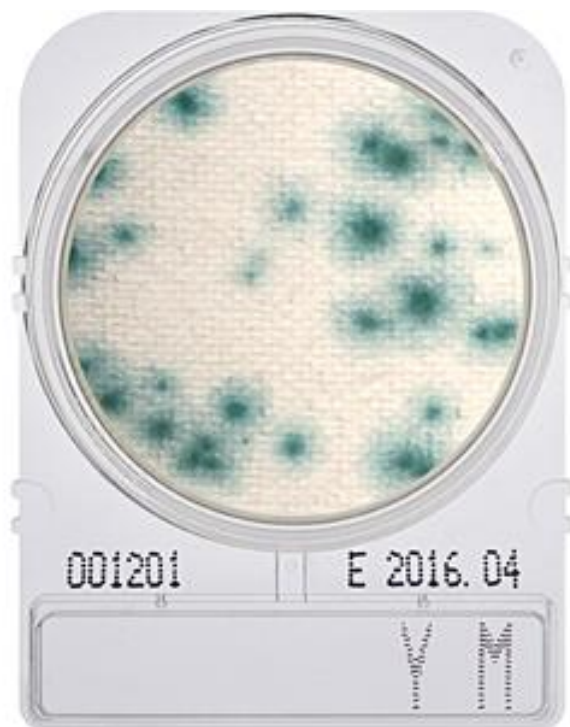
- ▶ Space: Sterile
- ▶ Cost: >\$9.00
- ▶ Time
- ▶ Skill
- ▶ Equipment
- ▶ Not field-friendly
 - ▶ Del Agua Kit

PROMISING LOW- COST ALTERNATIVES

CompactDry

- ▶ Data output: *E. coli* colony count
- ▶ Assumption for all tests: equipment and practices are sterile
- ▶ Step 1: Remove lid from CompactDry dry tray well
- ▶ Step 2: Inoculate by pipetting 1 mL of sample onto center of dry tray well
 - ▶ Do not touch pipette tip to matrix
- ▶ Step 3: Replace lid and label
 - ▶ Write dilution
- ▶ Step 4: Incubate at 35C for 24 hours

CompactDry



CompactDry

PROS

- ▶ Cost per test: \$1.50-\$5.00
- ▶ Time per test: 2-10 minutes
- ▶ Lab requirement: no
- ▶ Skilled Personnel: no
- ▶ Ease of learning: 5/5
- ▶ Ease of use: 4/5

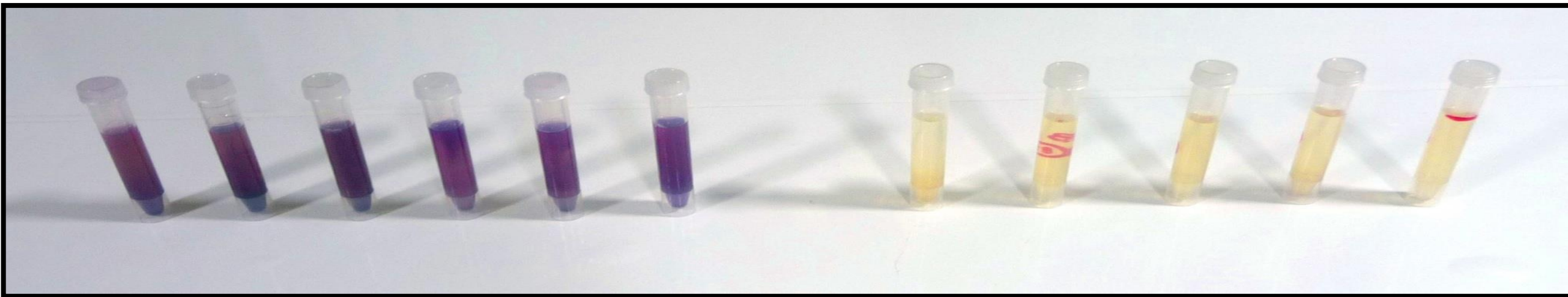
CONS

- ▶ Incubation: yes

LOW-COST PRESENCE-ABSENCE (LCPA)

- ▶ Data output: Presence-Absence (Detection Limit: 10CFU/100mL)
- ▶ Step 1: Put 0.31g of M-Tec or Aquatest (open source broth media) in 10mL vial
 - ▶ If testing drinking water samples, add 1 tablets of Sodium Thiosulfate
- ▶ Step 2: Pour 10mL of sample into vial
- ▶ Step 3: Shake to dissolve completely
- ▶ Step 4: Incubate at 35C for 24 hours
 - ▶ At ambient temp (25C or above) incubation time is 48hours

LOW-COST PRESENCE-ABSENCE (LCPA)



LOW-COST PRESENCE-ABSENCE (LCPA)

PROS

- ▶ Cost per test: \$0.50 (going down further)
- ▶ Time per test: 1-2 minutes
- ▶ Lab requirement: no
- ▶ Incubation: no
- ▶ Skilled Personnel: no
- ▶ Ease of learning: 5/5
- ▶ Ease of use: 5/5

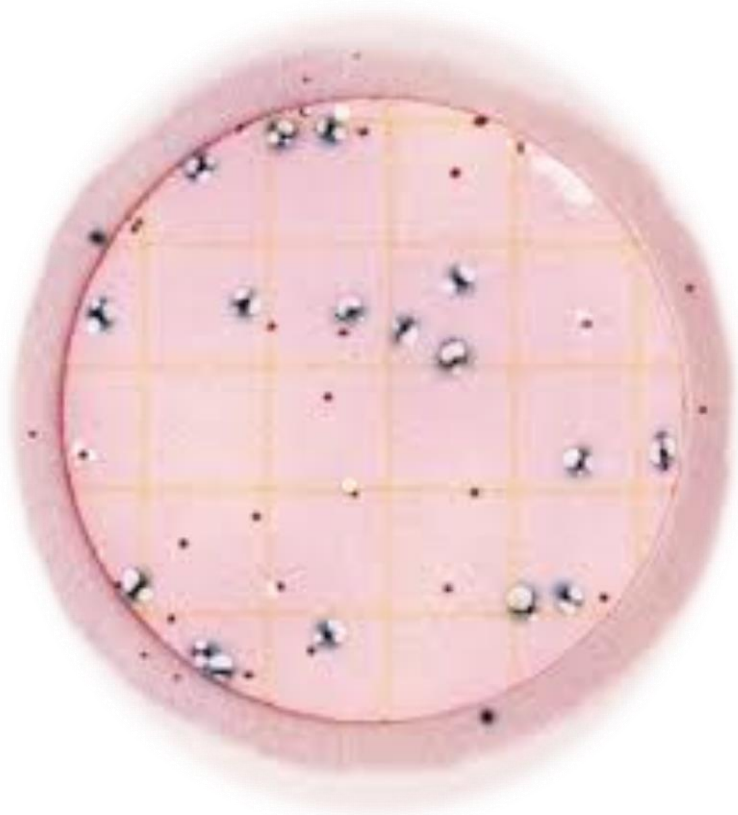
CONS

- ▶ Cannot differentiate between “low-risk” and “safe”.

3M Petrifilm

- ▶ Data output: *E. coli* colony count
- ▶ Step 1: Lift top film and pipette 1 mL of sample onto center of bottom film
- ▶ Step 2: Roll top film down onto sample gently to prevent pushing sample off film and to avoid trapping any air bubbles
- ▶ Step 3: Distribute sample across gel by applying pressure
- ▶ Step 4: Let gel dry for 1 minute and incubate at 35C for 24 hours

3M Petrifilm



3M Petrifilm

PROS

- ▶ Cost per test: \$1.30-\$5.00
- ▶ Time per test: 2-10 minutes
- ▶ Lab requirement: no
- ▶ Skilled Personnel: no
- ▶ Ease of learning: 4/5
- ▶ Ease of use: 3/5

CONS

- ▶ Incubation: yes
- ▶ Prone to bubble formation

Colitag

- ▶ Data output: Presence/Absence
- ▶ Step 1: Add powdered media packet to 100 mL water sample
- ▶ Step 2: Shake (minimum 25x) to dissolve
- ▶ Step 3: Incubate at 35C for 24 hours

Colitag



- Observe for yellow color and fluorescence under UV light (yellow and fluorescence = positive for *E. coli*)

Colitag

PROS

- ▶ Time per test: 2-3 minutes
- ▶ Lab requirement: no
- ▶ Skilled Personnel: no
- ▶ Ease of learning: 5/5
- ▶ Ease of use: 4/5
- ▶ Detection limit is 1CFU/100mL

CONS

- ▶ Cost per test: $\geq \$4.50$
- ▶ Incubation: yes
- ▶ Only detects “safe” vs “unsafe”

Compartment Bag Test

- ▶ Data output: Presence/Absence and MPN
- ▶ Step 1: Mix chromogenic *E. coli* broth culture medium with 100 mL of the water sample and dissolve for 15 minutes
- ▶ Step 2: Pour into the compartment bag
- ▶ Step 3: Gently squeeze the bag to distribute the correct volumes into the designated compartments
- ▶ Step 4: Seal bag with a reusable plastic clip and incubate at 35C for 24 hours or room temperature for 48 hours

Compartment Bag Test



- ▶ *E. coli* detected through production of blue-green color change in water sample

Compartment Bag Test MPN

Compartment #					MPN/100mL	Upper 95% Confidence Interval/100mL	Health Risk Category Based on MPN and Confidence Interval
1	2	3	4	5			
10mL	30mL	56mL	3mL	1mL			
					0.0	2.87	Low Risk/Safe
					1.0	5.14	Intermediate Risk/ Probably Safe
					1.0	4.74	
					1.1	5.16	
					1.2	5.64	
					1.5	7.81	
					2.0	6.32	
					2.1	6.85	
					2.1	6.64	
					2.4	7.81	
					2.4	8.12	
					2.6	8.51	
					3.2	8.38	
					3.7	9.70	
					3.1	11.36	Intermediate Risk/ Possibly Safe
					3.2	11.82	
					3.4	12.53	
					3.9	10.43	
					4.0	10.94	
					4.7	22.75	
					5.2	14.73	
					5.4	12.93	
					5.6	17.14	
					5.8	16.87	
					8.4	21.19	
					9.1	37.04	
					9.6	37.68	
					13.6	83.06	High Risk/Possibly Unsafe
					17.1	56.35	High Risk/Probably Unsafe
					32.6	145.55	
					48.3	351.91	High Risk/Probably Unsafe
					>100	9435.10	Unsafe

Compartment Bag Test

PROS

- ▶ Cost per test: unclear
- ▶ Lab requirement: no
- ▶ Incubation: no
- ▶ Skilled Personnel: no
- ▶ Ease of learning: 3.5/5
- ▶ Ease of use: 3.5/5

CONS

- ▶ Time per test: 17-20 minutes
- ▶ Filling compartments to correct volumes can be tricky

SUMMARY

Summary

Name of Test	Type	Cost/sample (US \$)	Field Processing Time (mins)	Lab Requirement	Incubation	Skilled Personnel	Ease of Learning	Ease of Use	Time to Result (hrs)
Compact Dry	Colony Count	1.5-5.0	2-10	No	Yes	No	5/5	4/5	24
LCPA	Presence/ Absence	0.50 (and decreasing)	1-2	No	Yes/No	No	5/5	5/5	24/48
3M Petrifilm	Colony Count	1.3-5.0	2-10	No	Yes	No	4/5	3/5	24
Colitag	Presence/ Absence	≥4.5	2-3	No	Yes	No	5/5	4/5	24
CBT	Most Probable Number	unknown	17-20	No	Yes/No	No	3.5/5	3.5/5	24/48

ACTION PLAN

Fecha	Actividad	Insumos	Responsable
Lunes 21.03.2016 Hrs. 09:00 – 10:20 Hrs. 10:20 – 11:20 Hrs. 11:20 – 11:30 Hrs. 11:30 – 12:20	Sesión teórica: 1) Técnica de análisis de coliformes fecales (Joe Brown y estudiantes) 2) Técnicas de análisis de protozoos, virus y bacterias (Aaron Bivins) 3) REFRIGERIO 4) Evaluación de condiciones biológicas utilizando macro invertebrados bentónicos (Giovanna Rocabado)	- Data show - Computador - Sala de reuniones - Refrigerio	- UNICEF (Alejandro)
Martes 22.03.2016 Hrs. 09:00 – 16:30	Trabajo de campo día 1 - Monitoreo del río Choqueyapu	- Equipos/patógenos - Equipos/macrobioinvertebrados - Movilidad A/6p (UNICEF) - Movilidad B/6p (VRHR)	- Georgia Tech (Joe Brown) - VRHR (Giovanna) - UNICEF (Alejandro) - VRHR (José Manuel)
Miércoles 23.03.2016 Hrs. 09:30 – 16:30	Trabajo de campo día 2 - Monitoreo del río Seco	- Equipos/patógenos - Equipos/macrobioinvertebrados - Movilidad A/6p (UNICEF) - Movilidad B/6p (VRHR)	- Georgia Tech (Joe Brown) - VRHR (Giovanna) - UNICEF (Alejandro) - VRHR (José Manuel)
Jueves 24.03.2016 Hrs. 09:00 – 12:30	Sesión teórica 1) Evaluación Coliformes f. (Joe) 2) Evaluación parásitos (Aaron) 3) Evaluación Cond. biológicas (Giovanna)	- Data show - Computador - Sala de reuniones - REFRIGERIO	- UNICEF (Alejandro)

VOLUNTEER